Sampling Protocol for Phosphine Application Monitoring

December 3, 2008

Prepared by:

Staff
In the
Special Purpose Monitoring Section

Signatures:

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Air Quality Surveillance Branch
Air Resources Board

The following protocol has been reviewed and approved by staff of the Air Resources Board (ARB). Approval of this protocol does not necessarily reflect the views and policies of the ARB, nor does the mention of trade names or commercial products constitute endorsement or recommendation for use.
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APPENDIX A: STANDARD OPERATING PROCEDURE ANALYSIS FOR PHOSPHINE

APPENDIX B: USE INFORMATION AND AIR MONITORING RECOMMENDATIONS FOR THE PESTICIDE ACTIVE INGREDIENT PHOSPHINE
1.0 Introduction

At the request of the California Department of Pesticide Regulation (DPR), (January 4, 2008 Memorandum, Warmerdam to Goldstene) the Air Resources Board (ARB) staff will monitor one application site for phosphine. This application monitoring study will be performed prior to, during and after an application of phosphine. Phosphine application monitoring is requested by DPR to fulfill the requirements of AB 1807/3219 (Food and Agricultural Code, Division 7, Chapter 3, Article 1.5, Section 14022(c)) which requires the ARB "to document the level of airborne emissions.... of pesticides which may be determined to pose a present or potential hazard..." when requested by the DPR. Monitoring is being conducted to coincide with the use of phosphine as a selective commodity fumigant.

The draft laboratory analysis method titled “Standard Operating Procedure Sampling and Analysis of Phosphine” dated November 2008, is included as Appendix A.

2.0 Project Goals and Objectives

The goal of this monitoring project is to collect and measure phosphine concentrations in ambient air prior to, during, and after an application. In addition, air from the chamber (prior to aeration) will also be collected and measured.

To achieve the project goal, the following objectives should be met:

1. Identify a suitable monitoring site that satisfies DPR’s air monitoring recommendations for phosphine.

2. Appropriate application of sampling/monitoring equipment to determine ambient phosphine concentrations.

3. Application of relevant quality assurance/quality control practices to ensure the integrity of field samples.

4. At the conclusion of the project, MLD will provide DPR with a final report containing all relevant information and data for this project.
3.0 Contacts

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4.0 Study Location and Design

Phosphine is used as a postharvest commodity fumigant in chambers or other enclosures. The location chosen for this study is a commercial commodity fumigation facility located near Ballico, CA. Monitoring will occur December, the month with the highest potential phosphine use.

Application Monitoring

Phosphine is a colorless and odorless gas, and is applied to postharvest commodities either as a gas or a metallic phosphide compound. For this study, the phosphine application will utilize a metallic phosphide (aluminum phosphide). A measured quantity (mass) of aluminum phosphide pellets will be placed inside a large chamber, and the chamber sealed. Humidity from the air inside the chamber reacts with the aluminum phosphide pellets creating phosphine gas.

The sampling/analysis method developed by the ARB Northern Laboratory Branch Special Analysis Section utilizes Silco canisters (Appendix A). During this study, a new canister sampler will be used (Tisch TE-323), enabling field staff to program equipment for unattended start and stop activation. The sampler can accommodate up to three (3) canisters for unattended sequential sampling. Canisters can be filled up to one (1) atmosphere above ambient. The target final canister pressure will be 10 psig.

Samples will be collected by pressurizing ambient air into a Silco canister. Approximately 3 lpm of air is pulled through the Tisch TE-323 inlet. By adjusting a turn style valve, a regulated portion of the 3 lpm air flow is forced into the sample canister. Except for the chamber sampler, the inlet heights will be placed at approximately 1.5 meters above the ground. The chamber sampler inlet will be placed approximately 1.0 meter above the floor.

Ten (10) Tisch TE-323 samplers were purchased for this study to reduce field staff exposure to phosphine gas during the fumigation and aeration process. Because the Tisch sampler can only be configured to sample three (3) canisters with each setup, the study will be divided into three (3) sampling episodes: Background, Fumigation & Aeration.

Background sampling: Four (4) primary samples, two (2) collocated samples and four (4) field spike samples will be deployed prior to phosphine fumigation. The four (4) primary samplers will be placed approximately 10 – 20 feet away from each side of the building. One (1) field spike sampler will be placed in parallel with each primary sampler. The collocated samplers will be placed at opposite sides of the building, one (1) of which is located at the expected downwind location. Sampling will occur concurrently. Sample duration will be configured for approximately 24 hour period. One (1) trip spike and one (1) trip blank will accompany the background samples to the field and back to the Laboratory. Please refer to Figure 5 for outside background sampling diagram.
Fumigation sampling: The fumigation process lasts between 24 hours to six (6) days, dependent on the expected phosphine concentration and ambient temperature. The duration of the fumigation process will be known prior to the application. As mentioned in the background sampling section above, one (1) trip spike and one (1) trip blank will accompany the samples to and from the field.

a) Inside chamber sample: A pre-programmed sampler will be placed inside the chamber and configured to automatically sample three (3) consecutive canisters. The sampler will be programmed so the last canister will complete sampling approximately one (1) hour before aeration begins. Due to the Tisch sampler design all three canisters must have the same flow set, and therefore the same sample duration. The inside chamber sample duration will be four (4) hours each.

b) Outside fumigation sampling: Outside the chamber, samplers will be placed 10 – 20 feet away from each of the four (4) building sides and 20 – 30 feet away from each of the four (4) building corners. A collocated sampler will be placed at the estimated downwind location. The samplers will be configured to fill each canister consecutively, with a sampling duration of four (4) hours each. Samplers will be programmed to complete sampling of the last canister sample approximately four (4) hours prior to the aeration process (allowing staff time to reconfigure). Please refer to Figure 5 for outside fumigation sampling diagram.

Aeration sampling: Pending local weather conditions, there are two (2) aeration monitoring scenarios. Similar to the background and fumigation sampling section above, one (1) trip spike and one (1) trip blank will accompany the aeration samples to and from the field.

a) Less than 5 knot/hour wind conditions: If local hourly averaged wind speeds are expected to be primarily less than 5 knots, then all four (4) corner samplers will be moved further outward between 40 – 60 feet away from the building, and the four (4) side samplers will be moved outward between 20 - 30 feet away from the building. Please refer to Figure 5 for this sampling scenario.

b) Greater than 5 knot/hour wind conditions: If the local hourly averaged wind speeds are expected to be primarily greater than 5 knots, all nine (9) samplers will be relocated at the expected downwind side of the building. Three (3) samplers will be located 20 – 30 feet away from the downwind corners and side of the building. Three (3) samplers will be located 60 – 70 feet away from the downwind corners and side of the building. Two (2) samplers will be located approximately 100 feet away from each downwind building corner. One (1) collocated sampler will be located next to the closest and most predominately downwind sampler. The samplers will be programmed to collect four (4) hour samples for each canister. Please refer to Figure 6 for the greater than 5 knot/hour wind speed sampling scenario.

An additional field stability spike will travel to the field along with the background canisters and return to the Laboratory with the aeration samples.

Every attempt will be made to shield all sampled canisters from direct sunlight to help reduce sampled phosphine losses. When possible, sampled canisters will be removed from the samplers and stored in a cool shaded location until they can be...
transported back to the Laboratory in Sacramento. Transportation of sampled canisters to Sacramento will occur as often as feasible during regular working hours.

DPR’s “Use Information and Air Monitoring Recommendations for the Pesticide Active Ingredient Phosphine”, dated September 2008 is included as Appendix B.

TABLE 1: Guidelines For Application Sampling Schedule

<table>
<thead>
<tr>
<th>Sample period begins:</th>
<th>Sample duration time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Background (pre-application)</td>
<td>10 canisters (total) – 24 hours each</td>
</tr>
<tr>
<td>Fumigation (inside building)</td>
<td>3 canisters (total) – 4 hours each.</td>
</tr>
<tr>
<td>Fumigation (outside building)</td>
<td>27 canisters (total) – 4 hours each</td>
</tr>
<tr>
<td>Aeration</td>
<td>27 canisters (total) – 4 hours each</td>
</tr>
</tbody>
</table>

Background air sampling will be completed approximately four (4) hours prior to the application. Fumigation sampling will be completed approximately four (4) hours prior to the aeration process. Aeration sampling will be completed after the third four (4) hour aeration sample has ended.

Eight samplers will be positioned around the application building perimeter. One sampler will be located at approximately the midpoint of each side of the building and one at each corner. A ninth sampler will be collocated at the expected downwind side or corner. Four (4) field spike samples will be collected during background sampling using the same ambient monitoring procedures (e.g., air flow rates, sample transportation and storage), and will be located in parallel to the ambient air samplers. Three (3) trip spikes, one (1) field stability spike and three (3) trip blanks will be included. (1 trip spike and 1 trip blank for the each sampling period, 1 stability spike spanning the entire project sampling period).
5.0 Sampling and Analysis Procedures

Special Purpose Monitoring Section (SPM) personnel will transport cleaned and evacuated canisters from MLD’s laboratory in Sacramento, to and from the sampling location and then return the sampled canisters back to MLD’s Sacramento laboratory. These samples will not be exposed to extreme conditions or subjected to rough handling that might affect sample integrity.

Prior to removing each sampled canister, the operator will assure that the canister valve is securely closed and the corresponding sample paperwork is complete. The collected canisters will be stored in a cool shaded location until they can be transported back the Laboratory. When received by the Laboratory, the canister samples will be analyzed as soon as possible.

All reported sampling times, including meteorological data, will be reported in Pacific Standard Time (PST).

The Northern Laboratory Branch (NLB) will supply SPM with cleaned and evacuated Silco canisters. NLB will perform analyses necessary to measure for phosphine concentrations in the sampled canisters and report results to SPM.

Laboratory analyses will be performed in accordance with applicable standard operating procedures (Standard Operating Procedure Sampling and Analysis of Phosphine) in Appendix A.
The following Silco canister validation and analytical quality control criteria should be followed during pesticide analysis.

1. **Sample Hold Time**: Sample hold time criteria will be established by the Laboratory. Samples not analyzed within the established hold time will be invalidated by the Laboratory.

2. **Duplicate Analysis**: Laboratory to establish relative percent difference (RPD) criteria for duplicate analysis. Lab to provide duplicate analytical results and RPD.

3. **Method Detection Limit (MDL)**: MDL sample analytical results less than the MDL shall be reported as a less than numerical value. This less than numerical value shall incorporate any dilutions/concentrations.

4. **Analytical Linear Range**: Any analytical result greater than the highest calibration standard shall be reanalyzed within the calibrated linear range.
### 6.0 List of Field Equipment

<table>
<thead>
<tr>
<th>Quantity</th>
<th>Item Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Met-One Auto met portable meteorology system having calibrated sensors to measuring 1 minute averages for wind speed, direction, ambient temperature, and relative humidity w/built-in data logger.</td>
</tr>
<tr>
<td>1</td>
<td>Measuring Wheel.</td>
</tr>
<tr>
<td>1</td>
<td>A 200 ft measuring tape.</td>
</tr>
<tr>
<td>1</td>
<td>Tripod and compass.</td>
</tr>
<tr>
<td>1</td>
<td>Global Positioning System (GPS) with backup batteries and carrying case.</td>
</tr>
<tr>
<td>1</td>
<td>Digital Camera with backup batteries and carrying case.</td>
</tr>
<tr>
<td>2</td>
<td>Alborg mass flow meter 0-100 cc/min.</td>
</tr>
<tr>
<td>10</td>
<td>Tisch TE-323 canister samplers.</td>
</tr>
<tr>
<td>79</td>
<td>Silco canisters (10 background, 30 fumigation, 27 aeration, 3 field spikes, 1 stability spike, 3 trip spikes, 3 trip blanks and 2 spares).</td>
</tr>
<tr>
<td>9</td>
<td>Tripods.</td>
</tr>
<tr>
<td>50</td>
<td>Batteries.</td>
</tr>
<tr>
<td>1</td>
<td>Hard hat for each individual.</td>
</tr>
<tr>
<td>1</td>
<td>Box of laboratory quality gloves</td>
</tr>
</tbody>
</table>
Figure 1: Chamber/Building Rear View
Figure 2: East Side View of Building
**Figure 3:** Site Map
Figure 4: Site Map (zoom)
**Figure 5:** Sample Data Sheet

**Figure 5:** Fumigation Monitoring (2X distance for aeration monitoring if WS < 5 knots)
Figure 6: Aeration Monitoring (if wind speeds > 5 knots)
CALIFORNIA AIR RESOURCES BOARD  
Pesticides  
SILCO Canister Pesticide Data/Sample Tracking Sheet

<table>
<thead>
<tr>
<th>Set-Up</th>
<th>Start</th>
<th>Stop</th>
</tr>
</thead>
<tbody>
<tr>
<td>Date</td>
<td>Time (PST)</td>
<td>LAB</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Time Vacuum MFC</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pressure or Vacuum</td>
</tr>
<tr>
<td>MFC Reading</td>
</tr>
<tr>
<td>Vacuum</td>
</tr>
</tbody>
</table>

** = Calibrated Guage Pressure or Vacuum

** = Calibrated Guage Pressure or Vacuum

### Sample Tracking

<table>
<thead>
<tr>
<th>Action</th>
<th>Transfer Method (Check one)</th>
<th>Name &amp; Initials</th>
<th>Date/Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Released by Lab</td>
<td>Carrier</td>
<td>Person</td>
<td></td>
</tr>
<tr>
<td>Received by Field</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Released by Field</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Received by Lab</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

---FOR LABORATORY USE ONLY---

Lab Comments:

---

Figure 7: Sample Data Sheet
7.0 Quality Control

Quality control procedures will be observed to ensure the integrity of samples collected in the field. National Institute of Standards and Technology (NIST) traceable transfer standards will be used to calibrate meteorological sensors and measure sample flow rates.

The metrological sensors will be calibrated and aligned following the procedures outlined in the standard operating procedures on the Air Monitoring Web Manual at the following link:

http://arb.ca.gov/airwebmanual/amwmn.php?c=5&t=sop

Each Silco canister will be assigned a field sample number that provides for identification of site, sample ID number, operator, and sample information as well as sample transfer information.

Field Spike (FS): A field spike will be prepared by the laboratory by injecting a known concentration of phosphine gas into a cleaned and evacuated Silco canister. The background field spikes (4 total) will be positioned in parallel with the primary background samples. The field spikes will be removed and handled identically to the other samples.

Field Stability Spike (SS): A field stability spike will be prepared by the laboratory by injecting a known concentration of phosphine gas into a cleaned and evacuated Silco canister. Similar to the Field Spike mentioned above, this spike will be created prior to background sampling and will remain in the field until it is brought in along with the last batch of samples. There will be only one (1) field stability spike during the monitoring portion of this study. The field stability spike will be removed and handled identically to the other samples.

Trip Spike (TS): A trip spike will be prepared by the laboratory by injecting a known concentration of phosphine gas into a cleaned and evacuated Silco canister at the same level as the field spike. The trip spike will be transported and analyzed along with the field spike. The trip spike is treated the same as a field spike with exception that it is not installed onto a sampler and not sampled.

Trip Blank (TB): A trip blank will be a cleaned and evacuated Silco canister transported to the field and returned to the Laboratory unopened.

Collocated (CO): A collocated (side-by-side) air sampler will operated exactly the same as the primary sampler and will be installed alongside the predominantly downwind sample site.
Site/Sample Identification

The phosphine application sampling sites will be named accordingly for the background, fumigation and aeration as follows:

Site Naming Examples:

N-BK = Northside background
W-BK-CO = collocated west side Background
E-F-1 = East side fumigation
S-A-3 = South side aeration

Letter Abbreviations as follows
N = North Side
W = West Side
E = East Side
S = South Side
NEC = NE Corner Sample
SEC = SE Corner Sample
SWC = SW Corner Sample
NWC = NW Corner Sample

Following the quality control procedures listed above will ensure the quality and integrity of the samples collected in the field and will ensure accurate field and lab data collection.
8.0 Deliverables

8.1 Air Quality Surveillance Branch Deliverables

Within 90 days from receipt of the final results report from the Northern Laboratory Branch (NLB), AQSB will provide DPR with a report containing the following topics:

1) Sampling Protocol.
2) Personnel Contact List.
3) Site Maps.
4) Site Photographs.
5) Site Descriptions and Measurements: Site, sampler, GPS coordinates, inlet height, distance to roads, site-specific comments, and amount of aluminum phosphide used.
6) The distance and direction of the sampler to the building.
7) A map of the monitoring site locations.
8) Sample Summary Table.
9) Field Sample Log.
10) Laboratory Analysis Reports with calculations in electronic format.
11) Meteorological data and supporting documentation.
12) Transfer Standards' Certification Reports.
13) Disk containing 1-minute averaged Meteorological Data.

In addition, the Special Purpose Monitoring Section (SPM) will prepare a project binder containing the above information. This binder will remain with SPM though available for viewing and review as requested.
8.2 Northern Laboratory Branch (NLB) Deliverables

Within 90 days from the last day of analysis, The NLB will provide SPM with a report that will include the following topics:

1) Table(s) of sample results to include:
   a. Sample identification (name).
   b. Date sample received from field.
   c. Date sample analyzed.
   d. Dilution ratio.
   e. Analytical results.

2) All equations used in calculating analytical results.

3) Table of duplicate results including calculated relative percent difference (RPD).

4) Table of collocated results.

5) Table of analytical results from all field, trip and laboratory spikes including percent recoveries.

6) Table of analytical results from all trip blanks.

7) Table of analytical results from all laboratory blanks, standards and control checks performed, including dates performed and relative percent recoveries if applicable.

8) Copy or location of analytical method or Standard Operating Procedures (SOP) used for analysis.

9) Section or provision listing or reporting any and all deviations from analytical SOP and this protocol.
APPENDIX A:
Standard Operating Procedure and Analyses for Phosphine

The Special Analysis Laboratory Section of MLD’s Northern Laboratory Branch will perform the analyses for phosphine collected by Silco canister method. This analytical procedure is entitled, Standard Operating Procedure Sampling and Analysis of Phosphine.
Standard Operating Procedure
Sampling and Analysis of Phosphine

Special Analysis Section
Northern Laboratory Branch
Monitoring and Laboratory Division

November 2008
Version 2

Approved by:
Russell Grace, Manager
Special Analysis Section
1. **SCOPE**

This method is for the sampling and analysis of phosphine in air samples using a six-liter Silco™ canister for sample collection. Collected samples are analyzed by gas chromatography using an automated cryogenic sampler.

2. **SUMMARY OF METHOD**

Air samples are collected in evacuated six-liter Silco™ canisters. The samples are collected automatically using a Tisch Environmental automatic sample collection system. Final pressures after collection are greater than ambient pressures. After collection, samples are analyzed using a Wasson ECE Instrumentation cryogenic sample concentrator and an Agilent 7890A gas chromatograph equipped with a flame photometric detector in the phosphorus mode. Confirmation of positive results is determined using an Agilent 5973 GC/MSD operated in the single ion monitoring mode (SIM). Sample analysis and quantitation uses an external standard method for instrument calibration. The estimated quantitation level (EQL) for this method is approximately 8.0 micrograms per cubic meter (µg/m³) prior to any sample dilution.

3. **INTERFERENCES / LIMITATIONS**

Method interference may be caused by contaminants in the Silco™ canisters or the Tisch sampler that can lead to discrete artifacts or elevated baselines. Analysis of samples containing high concentrations of early eluting components may cause significant contamination of the analytical equipment. A system blank must be analyzed with each batch of samples to detect any possible method or instrument interference.

4. **EQUIPMENT AND CONDITIONS**

   A. Instrumentation:
      Agilent Instruments 7890A gas chromatograph (GC) with a flame photometric detector (FPD) equipped with a phosphorus filter.

      GC Column:
      J&W GS Gaspro 30 meters by 0.32 millimeter ID (or equivalent)

      GC Temperature Program:
      Initial temperature 0 degrees centigrade (° C) for 5 minutes
      0 to 60 ° C at 5° C/min
      60 to 100° C at 50 ° C/min

      GC Inlet Parameters:
      Pressure at 12.78 pounds per square inch (psi)
      Flow at 2.0 ml/min
      GC Detector FPD:
Heat at 225 °C
Hydrogen Flow at 75 ml/min
Air Flow at 100 ml/min
Makeup Flow at 58 ml/min

Wasson ECE cryogenic sample concentrator with Nafion dryer:
Cryo Temp #1 at -140º C
Cryo Temp #2 at -150º C
Sample Oven at 200 º C
Transferline Temp at 150º C
Mass Flow at 35 ml/min
Line Purge Time 30 seconds

Agilent 5973 mass selective detector (MSD) with a 6890 GC
Acquisition Mode: SIM
Tune File: PFTBA autotune
Ions monitored: 31.0, 33.0, 34.0
Quant Ion: 34.0

B. Auxiliary Apparatus
   Restek six liter Silco™ canisters with Silco™ valves

C. Reagents
   Phosphine gas at 10 ppm +/-2% H.P. Gas Products, Inc.

D. Gases
   Helium, grade 5 or better
   Liquid Nitrogen at 22 pounds per square inch (psi)
   Nitrogen, grade 5
   Compressed air, ultra zero
   Hydrogen, supplied by a Whatman Hydrogen generator

5. SAMPLE COLLECTION

A. Samples are collected using a Tisch Environmental automated sampler set to deliver ambient air over a fixed amount of time (3 to 12 hours depending on sampler location).
B. Six liter canisters will be filled so the ending pressure will be above ambient in the range of 10 to 20 psig (psi gauge).
C. Phosphine is stable for at least five days when kept at ambient temperatures. See section 8F for storage stability summary.
6. ANALYSIS OF SAMPLES

a) Connect each canister to a port on the Wasson ECE cryoconcentrator using a short length of polypropylene tubing. Reserve ports one and two for the blank and calibration standard.

b) For this method the standard volume will be 100 milliliters.

c) Perform an initial calibration curve using the following volumes of known concentrations of Phosphine: 15, 25, 50, 75, 100 milliliters. At least five (5) points must be analyzed to establish a calibration curve. Appendix 1 lists the standard concentrations used when the EQL is approximately 8.0 µg/m³.

d) Prepare a sample sequence for the GC. The sequence should include a system blank and a continuing calibration verification standard (CCV) for every ten (10) samples analyzed. If this batch of samples includes a method blank and/or LCS, they should be run prior to field samples to verify that QC criteria have been met.

e) To minimize excessive carry over of contaminants from one analysis to the next, a system blank should be run after every twenty (20) samples or more frequently if indicated by sample chromatograms. In no case should a sample contaminant interfere with the peaks of interest. This will be verified by the absence of a peak in the analyte retention time window during the system blank analysis.

f) Review and edit the quantitation reports as needed.

g) Rerun samples with results greater than the EQL by GC/MSD to confirm the presence of phosphine.

h) Samples with concentration greater than the upper point of the calibration curve must be run at a smaller volume. Every attempt should be made to have the results fall within the upper half of the calibration curve. If running a volume of 15 mls results in a value greater than the upper calibration point, then the sample will need to be diluted with compressed nitrogen. Either add nitrogen to the original canister being sure to record the beginning and ending pressures, or transfer a known amount of sample from the original canister into a clean fully evacuated canister. Pressurize with nitrogen again recording the final pressure.

i) The final results will be adjusted by an appropriate dilution factor and reported in µg/m³.

j) The atmospheric concentration is calculated as follows:

\[ \text{Ambient Sample Conc. (µg/m³)} = \frac{\text{Sample Vol (ml)} \times \text{Instrument result (µg/m³)} \times \text{Dilution Factor}}{100 \text{ ml}} \]

k) Given instrument sensitivity and a minimum sample volume of 15 ml the EQL for this method will be approximately 8.0 µg/m³.
7. QUALITY ASSURANCE

A. Instrument Reproducibility

Establish the reproducibility of the instrument and analytical method as follows: Analyze three different volumes of standard (low, medium, and high levels) by injecting each five times. Table 1 lists the results for the Phosphine instrument reproducibility.

<table>
<thead>
<tr>
<th>Phosphine (ug/m³)</th>
<th>Low Level (15ml)</th>
<th>Medium Level (50ml)</th>
<th>High Level (100ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>7.74</td>
<td>25.9</td>
<td>51.1</td>
</tr>
<tr>
<td></td>
<td>7.69</td>
<td>25.9</td>
<td>58.3</td>
</tr>
<tr>
<td></td>
<td>7.75</td>
<td>25.4</td>
<td>54.2</td>
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<tr>
<td></td>
<td>8.26</td>
<td>27.2</td>
<td>52.6</td>
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<tr>
<td></td>
<td>7.79</td>
<td>26.9</td>
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<td></td>
<td>7.846</td>
<td>26.26</td>
<td>53.76</td>
</tr>
<tr>
<td></td>
<td>0.234</td>
<td>0.757</td>
<td>2.765</td>
</tr>
<tr>
<td></td>
<td>2.984</td>
<td>2.883</td>
<td>5.142</td>
</tr>
</tbody>
</table>

B. Linearity

A five point calibration is performed. Calibration standards ranging from at or near the EQL to approximately 7 times higher are used for phosphine. The results are used to calculate calibration curves using linear or quadratic regression. An $r^2$ of 0.995 or higher is required for an initial calibration to be acceptable. See Appendix 1 for an example calibration curve. A CCV will be run at the start of each analytical batch, and after every tenth sample to verify the system linearity. The CCV quantitated value must be within 25% of the actual value.

C. Method Detection Limit

Method detection limits (MDL) are based on the US EPA MDL calculation. Using the analysis of seven replicates of a low-level standard, the MDL and EQL for Phosphine are calculated as follows:

$$\text{MDL} = 3.143 \times \text{STD}$$
EQL = 5*MDL

STD equals the standard deviation of the calculated results for the seven replicate spikes. The calculated MDL for phosphine is 0.743 µg/m³ based on a 100 ml sample analysis volume. The EQL for phosphine based on the MDL would be approximately 4.0 µg/m³. Although statistically achievable, based on the daily standard concentration of approximately 50 µg/m³ and the smallest volume to be sampled, 15 ml, the actual lowest concentration analyzed will be approximately 8.0 µg/m³. Thus the EQL for reporting results will be approximately 8.0 µg/m³. Since the EQL used is based on the calibration curves high point, the MDL will also be effected by the same factor which in most case is approximately two. Thus the MDL used for this analyte will be around 1.5 µg/m³.

D. Laboratory Control Sample

A laboratory control sample (LCS) is included with each analytical batch. The LCS stock standard should come from a different source or lot than the daily calibration standards. The analytical value of the LCS must be within three standard deviations of its historical mean. If the LCS is outside these limits then the samples in the analytical batch must be reanalyzed. The LCS is prepared like the calibration standards using a volume of standard that will result in a final concentration that is in the middle of the calibration curve, in this case approximately 30 µg/m³.

E. Storage Stability

Storage stability was determined for phosphine by spiking an evacuated canister and collecting ambient air using the Tisch Environmental sampler. This sample was then analyzed on the following days: 0, 1, 5, 13, and 25 days. Table 2 lists the results for the storage stability study.

<table>
<thead>
<tr>
<th>Day</th>
<th>Sample 1 %recovery</th>
<th>Sample 2 %recovery</th>
<th>Sample 3 %recovery</th>
<th>Average %recovery</th>
<th>Standard Dev</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>87.17</td>
<td>89.52</td>
<td>100.31</td>
<td>92.34</td>
<td>7.008</td>
</tr>
<tr>
<td>1</td>
<td>89.40</td>
<td>97.85</td>
<td>94.12</td>
<td>93.79</td>
<td>4.235</td>
</tr>
<tr>
<td>5</td>
<td>92.94</td>
<td>93.29</td>
<td>94.52</td>
<td>93.58</td>
<td>0.828</td>
</tr>
<tr>
<td>13</td>
<td>60.30</td>
<td>62.30</td>
<td>61.02</td>
<td>61.21</td>
<td>1.013</td>
</tr>
<tr>
<td>25</td>
<td>48.43</td>
<td>49.12</td>
<td>47.82</td>
<td>48.46</td>
<td>0.650</td>
</tr>
</tbody>
</table>

Because there is a significant loss between day 5 and 13 all samples should be analyzed within 5 to 6 days of sampling.
H. Safety

This procedure does not address all of the safety concerns associated with chemical analysis. It is the responsibility of the analyst to establish appropriate safety and health practices. For hazard information and guidance refer to the material safety data sheets (MSDS) of any chemicals used in this procedure. Phosphine gas is highly toxic at levels greater than 11.5 mg/kg of body weight. All prep of standards and expected high samples should be performed in a shielded fume hood.
Appendix 1

Calibration Standard Preparation for Phosphine

The certified gas standard used for calibration was purchased from H.P. Gas Products, Inc., Baytown, Texas and has the following specification:

<table>
<thead>
<tr>
<th>Cylinder No:</th>
<th>CC177430</th>
</tr>
</thead>
<tbody>
<tr>
<td>Expiration date:</td>
<td>December 10, 2008</td>
</tr>
<tr>
<td>Phosphine:</td>
<td>10 ± 2 ppm</td>
</tr>
</tbody>
</table>

The calibration standard is made by taking an aliquot of the stock standard (10ppm) and diluting in a six liter Silco™ canister with nitrogen. A typical dilution is as follows:

75ml of Phosphine (10ppm) at 13,906 µg/m³

\[ \text{[ µg/m}^3\text{ = ppm x molecular weight x 40.90]} \]

Pressurize canister to approximately 30 psig

\[ \text{[ Volume in ml = (Final Pressure (psig)/14.7 psig x 6000ml) + 6000ml]} \]

Resulting concentration in can is approximately 57 µg/m³

Final concentration µg/m³ = vol of std/vol in canister x std conc (µg/m³)

A minimum of five sample volumes are used to generate the calibration curve, with the standard at 15 ml being the low point. The sample volumes for the calibration curve are 15, 25, 50, 75, 100 ml. The 100ml represents a concentration of 57 µg/m³. The low point (15ml) equates to approximately 8.5 µg/m³.

All continuing calibration verification standards (CCV) are run at 15 ml, while all samples are run at 100 ml.

Initial calibration curve acceptance requires an \( r^2 \) of at least 0.995.
APPENDIX B:

USE INFORMATION AND AIR MONITORING RECOMMENDATIONS FOR THE PESTICIDE
ACTIVE INGREDIENT PHOSPHINE

http://www.cdpr.ca.gov/docs/emon/pubs/tac/recomm/air_rpt_phosphine.pdf